

## Original Research Article

<https://doi.org/10.20546/ijcmas.2019.810.026>

## Allelopathic Potential Assessment of Sorghum and Sunflower on Germination Characteristics of *Phalaris minor* and Wheat

Arya Kumar Sarvadamana\*, V. Pratap Singh, S. K. Guru, S. P. Singh,  
Tej Pratap, Sirazuddin and Suprava Nath

Department of Agronomy, G. B. Pant University of Agriculture & Technology,  
Pantnagar, India

\*Corresponding author

### ABSTRACT

#### Keywords

Aqueous extract,  
Allelopathy,  
Germination  
percent, Sorghum,  
sunflower

#### Article Info

Accepted:  
04 September 2019  
Available Online:  
10 October 2019

A laboratory experiment was conducted to assess the allelopathic effect of aqueous extracts of sorghum and sunflower on germinating seedlings of *Phalaris minor* and wheat. 10% (w/v) aqueous extracts of sorghum and sunflower were applied on the seed of *Phalaris minor* and wheat and the parameters like germination percent, relative germination ratio, speed of germination, mean germination time, days to 50% germination were calculated. In all the calculated parameters it was found that the aqueous extracts of both sorghum and sunflower are highly allelopathic to *Phalaris minor* and they have a very little effect on wheat. Both sorghum and sunflower aqueous extracts significantly reduced the germination of *Phalaris minor* as compared to control (distilled water), but sunflower was more allelopathic than sorghum with 85.5% of germination inhibition.

### Introduction

Wheat is the most important cereal crop of the world and the second most important in India having 30.17 million hectare area and 99.70 million tonnes of production (Anonymous, 2018). It has a lion share of about 35% in national food basket. Wheat is a very rich source of nutrients, which have 76% carbohydrate, 1.5% fat and 13% protein. Albumins, globulins are the major proteins of the gluten complex. The contents of minerals

and of dietary fibers are very low; 0.5% and 1.5%, respectively (Belderok *et al.*, 2000). Rice-wheat cropping system is the most dominant cropping system of India. In Rice-Wheat cropping system along with other traditional weed flora *Phalaris minor* became a major problem causing drastic reduction in wheat yield (Chhokar *et al.*, 2006). Wheat fields in Northern India are badly infested with wide range of grassy and non grassy weeds in general and *Phalaris minor* Retz. in particular. Continuous application of herbicides at suboptimal dose,

*Phalaris minor* gradually developed resistance against these (Chhokar and Malik, 2002). Hence, to tackle this problem alternative weed control methods like allelopathic approach can be a potential tool in future.

### Materials and Methods

The laboratory experiment was conducted in seed physiology laboratory, Department of Agronomy, Collage of Agriculture, Govind Ballabh Pant University of Agriculture and Technology, Pantnagar, Udham Singh Nagar, Uttarakhand. Sorghum and sunflower biomass was collected from the Norman E. Borlaug Crop Research Centre, Govind Ballabh Pant University of Agriculture and Technology, Pantnagar and were shade dried for about one week at Weed management block (D2). Then the biomass of sorghum and sunflower shade dried at 65±5 °C for 72 hours and finely ground with an electric grinder after fully drying. The grinded biomass was placed in an air tight container in cool and dry place. The ground biomass was weighed using high precision electronic balance. After this the grinded biomass was well mixed in distilled water and soaked for 48 hours in refrigerator to avoid fermentation. This was then filtered using muslin cloth to get aqueous extract. Using this method aqueous extracts of sorghum and sunflower of 10% concentration (w/v, dry weight basis) was prepared.

The experiment was carried out in Completely Randomized Design (CRD), with three treatments *i.e.* control (distilled water), 10% aqueous extract of sorghum, 10% aqueous extract of sunflower, each with three replications.

Healthy, disease free and uniform seeds of both wheat (variety-DBW 17) and *Phalaris minor* were collected and treated with thiram to avoid fungal infection. Petridishes with 15cm diameter was also taken and cleaned

properly with chromic acid to remove all the dirt adhering with it. Then it was washed with distilled water and put in hot air oven for drying. After the drying is completed the petridishes were sterilized with methanol.

On the bottom side of petridish two filter papers were put to hold the aqueous extract. Then 50 seeds were arranged uniformly on the filter paper and 5ml aqueous extract was added to each petridish with help of pipette.

Finally the petridishes were put in a seed incubator at 25±1 °C for 7 days for the germination procedure to be completed.

The parameters were calculated as the formula given below-

$$\text{Germination percent (AOSA, 1998)} = \frac{\text{No. of seeds produced normal seedlings}}{\text{No. of seedlings set for germination}} \times 100$$

$$\text{Relative germination ratio (Rho and Kill, 1986)} = \frac{\text{germination percent of treated seeds}}{\text{germination percent of control}} \times 100$$

$$\text{Speed of germination (AOSA, 1983)} = \frac{\text{No. of seedlings}}{\text{Day of 1st count}} + \dots + \frac{\text{No. of seedlings}}{\text{Day of final count}}$$

$$\text{Mean germination time (Bonner, 1983)} = \frac{\sum(n \times d_i)}{\text{No. of seeds set for germination}}$$

Where,

n= number of seeds newly germinated on day di

di= i<sup>th</sup> day after incubation

$$\text{Days to 50\% germination (Dezfuli et al., 2008)} = t_i + \frac{\{(N/2) - n_i\} \times (t_i - t_j)}{(n_i - n_j)}$$

$$T50 = t_i + \frac{\{(N/2) - n_i\} \times (t_i - t_j)}{(n_i - n_j)}$$

Where,

N= final number of germinated seeds

$n_i$  and  $n_j$  are cumulative number of seeds germinated by adjacent counts at times  $t_i$  and  $t_j$ , where  $n_i < N/2 < n_j$

## Results and Discussion

### Germination per cent (7 days after incubation)

The germination per cent of wheat was found non significant between the treatments, which is a desirable character. But, numerically maximum germination percent was found in case of control and extract of sunflower (96.00%). However, minimum (94.0%) was achieved by the application of extract of sorghum at 10 % (Table 1).

The germination per cent *Phalaris minor* was found highest in control (82.66%). In case of aqueous extract of sunflower lowest germination percent of *Phalaris minor* was found (12%), which is significantly lower than both control and aqueous extract of sorghum at same concentration. The application of aqueous extract at 10 percent concentration reduced the population of *Phalaris minor* by 85.4% and 42.7% under sunflower and sorghum extracts respectively with respect to control.

It is evident that sorghum and sunflower extracts have strong allelopathic effect on *Phalaris minor* but little allelopathic effect on wheat. It also matches with the findings of Anjum and Bajwa (2005).

Among sorghum and sunflower, sunflower has higher inhibitory effect on *Phalaris minor* and it also does not affect germination of wheat, there

by fulfilling our aim. Hence, sunflower aqueous extract can be used as a potential tool for *Phalaris minor* control in wheat.

Based on this findings a field experiment was also conducted by the sequential (pre *fb* early post) application of both the aqueous extracts of sorghum and sunflower at same concentration (10%).

### Relative germination ratio

Relative germination ratio gives information about germination percent of all the treatments in comparison to control. In wheat relative germination ratio was found non-significant among the treatments.

It shows that all the treatments are alike with respect to their percentage of germination. This gives an idea that the allelochemicals present in sorghum and sunflower don't check germination of wheat seeds. But, while the *Phalaris minor* seeds lowest germination ratio was obtained with aqueous extract of sunflower treated seeds (14.5) followed by aqueous extract of sorghum treated seeds (57.16).

This type of result comes due to a little allelopathic effect of sorghum and sunflower on wheat and a very high allelopathic effect on *Phalaris minor* seeds. This type of result matches with the study of Bhadoria, (2011).

### Speed of germination

The aqueous extract of sorghum as well as sunflower had no effect on the speed of germination of wheat.

However, in case of *Phalaris minor* the minimum speed was recorded with application of aqueous extract of sunflower (10%), which was significantly lower than the aqueous extract of sorghum (10%) (Table 2).

**Table.1** Effect of aqueous extracts of sorghum and sunflower (10%) on germination percent and relative germination ratio of wheat and *Phalaris minor*

Treatment	Germination percent (%)		Relative germination ratio	
	Wheat	<i>Phalaris minor</i>	Wheat	<i>Phalaris minor</i>
Control (distilled water)	96.00	82.66	100	100
Sorghum (10%)	94.00	47.33	97.74	57.16
Sunflower (10%)	96.00	12.00	99.55	14.5
SEm±	0.94	1.30	1.50	1.35
CD (5%)	NS	4.50	NS	4.77

**Table.2** Effect of aqueous extracts of sorghum and sunflower (10%) on Speed of germination, Mean germination time and Days to 50% germination of wheat and *Phalaris minor*

Treatment	Speed of germination (no.of seedlings germinated/ day)		Mean germination time (days)		Days to 50% germination	
	Wheat	<i>Phalaris minor</i>	Wheat	<i>Phalaris minor</i>	Wheat	<i>Phalaris minor</i>
Control (distilled water)	6.9	5.9	5.34	3.36	3.12	4.25
Sorghum (10%)	6.73	3.38	5.22	1.89	3.17	4.31
Sunflower (10%)	6.83	1.57	5.47	0.54	3.13	4.29
SEm±	0.06	0.08	0.05	0.02	0.04	0.07
CD (5%)	NS	0.30	0.18	0.08	NS	NS

While both the aqueous extracts were significantly superior over control.

It is obvious that aqueous extract of sunflower (10%) has a inhibitory effect on the germination of *Phalaris minor*.

This finding are in agreement with the reports of Ghafar *et al.*, (2001), who stated that five allelochemicals like (chlorogenic, caffeic, syringic, vanillic and ferulic acid) in leaves, three allelochemicals in stem (chlorogenic, ferulic and vanillic acids) and only one (ferulic acid) in the roots have allelopathic effect on *Phalaris minor*.

### Mean germination time

The mean germination time of wheat in aqueous extract of sunflower 10% (5.47) is statistically at par with control (5.34), however it was significantly higher than aqueous extract of sorghum 10% (5.22). The mean germination time of *Phalaris minor* was found significantly lower by the application of both the aqueous extract of sorghum (1.89) and sunflower (0.54) than control (3.36).

This results reveals that sunflower has a lesser inhibitory effect on wheat followed by sorghum extract, and a very strong

allelopathic effect on *Phalaris minor*. Hence, it may be concluded that sunflower can be a potential source in managing *Phalaris minor* in wheat crop.

### Days to 50% germination (T<sub>50</sub>)

Days to 50% germination was found non significant over the treatments on both wheat and *Phalaris minor*. But numerically it was found higher in aqueous extract of sorghum 10% (4.31) on *Phalaris minor*. This might be due to the allelo-chemicals affect the germination process uniformly throughout the germination period.

### References

Anjum, T. and Bajwa, R. 2005 A bioactive annuionone from sunflower leaves. *Phytochem* 66: 1919–1921.

Anonymous. 2018. *Agricultural Statistics at a Glance-2016*. Directorate of Economics and Statistics, Ministry of Agriculture, Govt. Of India.

Association of Official Seed Analysis (AOSA) 1983. Seed vigor testing handbook, Contribution No. 32 to the Handbook of seed testing.

Association of Official Seed Analysis (AOSA) 1998. Rules of testing seeds. *Journal of Seed Technology*. 12: 1-112.

Belderok, B.; Mesdag, H. and Donner, D.A.

2000. Bread-Making Quality of Wheat. *Springer*, New York.

Bonner, F.T. 1983. Germination responses of loblolly pine to temperature differences on a two-day thermogradient plate. *J. Seed Technol.*, 8(1): 6-14.

Chhokar, R.S. and Malik, R.K. 2002. Isoproturon resistant *Phalaris minor* and its response to alternate herbicides. *Weed Technology*. 16: 116-123.

Chhokar, R.S.; Sharma, R.K.; Chauhan, D.S. and Mongia, A.D. 2006. Evaluation of herbicides against *Phalaris minor* in wheat in north western plains. *Weed Research*. 46: 40-49.

Dezfuli, P.M.; Sharif-zadeh, F. and Janmohannadi, M. 2008. Influence of priming techniques on seed germination behavior of maize inbred lines (*Zea mays* L.) *ARPN Journal of Agriculture and Biological Science*, 3(3): 22-25.

Ghafar, A.; Saleem, B.; Haq, A. and Qureshi, M.J. 2001. Isolation and identification of allelochemicals of sunflower (*Helianthus annuus* L.). *Int J Agric. Biol.*, 3: 21–22.

Rho, B.J. and Kill, B.S. 1986. Influence of phytotoxin from *Pinus rigida* on the selected plants. *Journal of Natural Science*. Wankwang university. 5:19-27.

### How to cite this article:

Arya kumar Sarvadamana, V. Pratap Singh, S. K. Guru, S. P. Singh, Tej Pratap, Sirazuddin and Suprava Nath. 2019. Allelopathic potential assessment of sorghum and sunflower on germination characteristics of *Phalaris minor* and wheat. *Int.J.Curr.Microbiol.App.Sci*. 8(10): 256-260. doi: <https://doi.org/10.20546/ijcmas.2019.810.026>